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## **On-line oxygen uptake rate and culture viability measurement of animal cell culture using microplates with integrated oxygen sensors**

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### **Abstract:**

O<sub>2</sub> uptake rates of animal cells (Chinese hamster ovary-CHO) were measured in 96-well microtiter plates by integrating with fluorescent sensors thereby measuring fluorescence intensity ratios of an O<sub>2</sub>-sensitive and an insensitive fluorophor. O<sub>2</sub> consumption rate was estimated from measured dissolved O<sub>2</sub> and from O<sub>2</sub> mass transfer coefficient determined in advance. Specific uptake decreased with time from  $3.2 \times 10^{-13}$  mol O<sub>2</sub> cell<sup>-1</sup> h<sup>-1</sup> at 15 h cultivation to  $1.8 \times 10^{-13}$  mol O<sub>2</sub> cell<sup>-1</sup> h<sup>-1</sup> at 48 h. Specific O<sub>2</sub> uptake was also determined by sampling from a spinner-flask culture giving identical values. A cell viability assay for cultures based on O<sub>2</sub> measurements is described in which cells are incubated outside the fluorescence reader and then the dissolved O<sub>2</sub> is measured only once at a fixed time after the start of incubation. This protocol can be directly applied for high-throughput measurements.

Key-words: CHO cells, culture viability, fluorescent oxygen sensor, microtiter plates, oxygen uptake rate